Protein Digestion and Quality of Goat and Cow Milk Infant Formula and Human Milk Under Simulated Infant Conditions

*Annet Maathuis, *Robert Havenaar, †Tao He, and *Susann Bellmann

ABSTRACT

Objective: The aim of this study was to determine the kinetics of true ileal protein digestion and digestible indispensable amino acid score (DIAAS) of a goat milk-based infant formula (GIF), a cow milk-based infant formula (CIF), and human milk (HM).

Methods: The GIF, CIF, and HM were investigated in an in vitro gastrointestinal model simulating infant conditions. Digested compounds were dialyzed from the intestinal compartment as bioaccessible fraction. Dialysate was collected in 15 to 60-minute periods for 4 hours. True ileal protein digestibility and DIAAS were determined as bioaccessible nitrogen (N) and amino acids.

Results: N bioaccessibility from the GIF showed similar kinetics to that of HM. The CIF showed a delay in N bioaccessibility versus the GIF and HM. In the 1st hour of digestion, N bioaccessibility was 19.9% ± 3.5% and 23.3% ± 1.3% for the GIF and HM, respectively, and 11.2% ± 0.6% for CIF (P < 0.05 vs HM). In the 3rd hour of digestion, the N bioaccessibility was higher (P < 0.05) for the CIF (28.9% ± 1.2%) than for the GIF (22.5% ± 1.6%) and HM (20.6% ± 1.0%). After 4 hours, the true ileal protein digestibility of the GIF, CIF, and HM was 78.3% ± 3.7%, 73.4% ± 2.7%, and 77.9% ± 4.1%, respectively. The DIAAS for the GIF, CIF, and HM for 0- to 6-month-old infants was 83%, 75%, and 77% for aromatic AA.

Conclusion: The protein quality is not different between the GIF, CIF, and HM, but the kinetics of protein digestion of the GIF is more comparable to that of HM than that of the CIF.

Key Words: DIAAS, digestible indispensable amino acid score, protein digestion kinetics, true ileal protein digestibility

What Is Known
- Growth of infants feeding goat or cow milk infant formula is not different.
- Goat, cow, and human milk differ in caseins.

What Is New
- True ileal protein digestibility of goat and cow milk infant formula is similar to that of human milk.
- Protein digestion of cow milk infant formula is delayed compared to goat milk infant formula and human milk.
- Digestible indispensable amino acid score can be determined in in vitro dynamic models simulating infant digestion.
- Digestible indispensable amino acid score of goat and cow milk infant formula for infants is not different compared to human milk.

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When breast-feeding is not sufficient or not possible, it is important to have high quality infant formulas available. Traditionally infant formula (IF) is based on cow milk proteins with an adapted ratio of casein to whey proteins by the addition of whey proteins. Nevertheless, there is an increasing consumer demand for goat milk-based IF (GIF). It has been shown that GIF provided growth and nutritional outcomes in infants similar to that of standard whey-dominant cow milk-based IF (CIF) (1,2). The knowledge, however, about protein digestion and availability for absorption of indispensable amino acids (IAAs) from IFs for infants is not yet completed. The Food and Agriculture Organization (FAO) (3) recommends to evaluate the protein quality based on true protein digestibility in the small intestine and the bioavailability of individual IAA. This should be used to calculate the Digestible Indispensable Amino Acid Score (DIAAS), which is the first limiting IAA as ratio of the IAA requirement for different age groups, for example, infants. The true IAA bioavailability should preferably be determined in humans (3). Digestion experiments in infants, however, have clinical and ethical drawbacks. Digestion experiments in young piglets as previously described (4,5) also have ethical constrains. An option is to use ex vivo gastrointestinal enzymes in static models (6,7). A better alternative might be the use of dynamic in vitro gastrointestinal models simulating infant digestion conditions (8,9). In the present study...
we used a dynamic in vitro gastrointestinal model (tiny-TIM) as described previously (10,11), improved with a 3-stage gastric compartment (12). This model has been validated for simulating the conditions in the upper gastrointestinal tract of neonates, infants, and toddlers (13), for studying the true ileal protein digestibility with an in vitro— in vivo correlation coefficient of 0.94 (10,11,14) and determining the in vitro DIAAS (11). Knowledge about IAA bioavailability of goat milk products is limited, but important for understanding GIFs. The aim of this study is to compare the kinetics of true ileal protein and AA digestibility and DIAAS of a GIF, a CIF, and HM in tiny-TIM under simulation of infant digestive conditions.

### METHODS

#### Products

In this study, we used a GIF and CIF manufactured under similar GMP processing by Ausnutria Hyproca (Zwolle, Netherlands), and HM pooled from 6 volunteers. The GIF and CIF were whey protein enhanced to a whey:casein ratio of 60:40. After signing an informed consent, HM was collected 3 to 5 months postpartum. To collect sufficient HM, it was necessary to freeze and store HM at −80°C. In daily practice, HM is also often stored under frozen conditions before use. HM was used in the experiments within 2 to 3 months after collection, after overnight thawing at 6°C.

Table 1. Analyzed concentration of nitrogen, indispensable amino acids (IAA) and dispensable amino acids (DAA) of reconstituted goat milk and cow milk based infant formula (IF) and human milk

<table>
<thead>
<tr>
<th></th>
<th>Goat milk-based IF mg/100 mL</th>
<th>%*</th>
<th>Cow milk-based IF mg/100 mL</th>
<th>%*</th>
<th>Human milk mg/100 mL</th>
<th>%*</th>
</tr>
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<tbody>
<tr>
<td>Nitrogen</td>
<td>276</td>
<td></td>
<td>262</td>
<td></td>
<td>161</td>
<td></td>
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<tr>
<td>Protein (N × 6.25)†</td>
<td>1725</td>
<td></td>
<td>1638</td>
<td></td>
<td>1006</td>
<td></td>
</tr>
<tr>
<td>Branched chain IAA</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Isoleucine</td>
<td>96.0</td>
<td>9.8</td>
<td>96.9</td>
<td>10.5</td>
<td>51.1</td>
<td>10.0</td>
</tr>
<tr>
<td>Leucine</td>
<td>175.4</td>
<td>17.9</td>
<td>173.9</td>
<td>18.8</td>
<td>96.3</td>
<td>18.9</td>
</tr>
<tr>
<td>Valine</td>
<td>112.9</td>
<td>11.5</td>
<td>105.8</td>
<td>11.4</td>
<td>58.8</td>
<td>11.5</td>
</tr>
<tr>
<td>Sulfur IAA</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Methionine</td>
<td>46.3</td>
<td>4.7</td>
<td>47.1</td>
<td>5.1</td>
<td>17.0</td>
<td>3.3</td>
</tr>
<tr>
<td>Cystine</td>
<td>34.1</td>
<td>3.5</td>
<td>29.8</td>
<td>3.2</td>
<td>23.1</td>
<td>4.5</td>
</tr>
<tr>
<td>Aromatic IAA</td>
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<tr>
<td>Phenylalanine</td>
<td>69.1</td>
<td>7.1</td>
<td>67.0</td>
<td>7.2</td>
<td>36.5</td>
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<tr>
<td>Tyrosine</td>
<td>60.2</td>
<td>6.1</td>
<td>63.6</td>
<td>6.9</td>
<td>40.5</td>
<td>7.9</td>
</tr>
<tr>
<td>Other IAA</td>
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<td></td>
<td></td>
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<tr>
<td>Lysine</td>
<td>157.5</td>
<td>16.1</td>
<td>136.4</td>
<td>14.7</td>
<td>64.8</td>
<td>12.7</td>
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<tr>
<td>Tryptophan</td>
<td>27.6</td>
<td>2.8</td>
<td>25.1</td>
<td>2.7</td>
<td>16.1</td>
<td>3.2</td>
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<tr>
<td>Threonine</td>
<td>113.8</td>
<td>11.6</td>
<td>99.7</td>
<td>10.8</td>
<td>45.6</td>
<td>8.9</td>
</tr>
<tr>
<td>Histidine</td>
<td>42.5</td>
<td>4.3</td>
<td>37.5</td>
<td>4.0</td>
<td>27.1</td>
<td>5.3</td>
</tr>
<tr>
<td>Arginine</td>
<td>44.7</td>
<td>4.6</td>
<td>43.6</td>
<td>4.7</td>
<td>32.7</td>
<td>6.4</td>
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<tr>
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<td></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Alanine</td>
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<td></td>
<td>68.6</td>
<td></td>
<td>35.2</td>
<td></td>
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<tr>
<td>Aspartic acid</td>
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<td></td>
<td>157.9</td>
<td></td>
<td>84.8</td>
<td></td>
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<tr>
<td>Serine</td>
<td>93.1</td>
<td></td>
<td>87.3</td>
<td></td>
<td>44.8</td>
<td></td>
</tr>
<tr>
<td>Glutamic acid</td>
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<td>316.8</td>
<td></td>
<td>179.5</td>
<td></td>
</tr>
<tr>
<td>Proline</td>
<td>137.7</td>
<td></td>
<td>135.0</td>
<td></td>
<td>86.0</td>
<td></td>
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<tr>
<td>Glycine</td>
<td>33.7</td>
<td></td>
<td>31.6</td>
<td></td>
<td>23.6</td>
<td></td>
</tr>
</tbody>
</table>

†DAA = dispensable amino acid; IAA = indispensable amino acid; IF = infant formula.

*Calculated as percentage of total IAA.

Not corrected for non-protein nitrogen.

The GIF (16.9 g) and CIF (18.9 g) powders were suspended in 112.5 mL water. For GIF, this was according to the label instruction; for CIF, it was 2 g higher than the label instruction to standardize the amount of protein intake at 2.1 g per experiment. The analyzed concentration of nitrogen (N) and AA in the reconstituted IFs and in the pooled HM (Table 1) was used to calculate the actual amount of protein and AA intake in the experiments.

#### Gastrointestinal Model

The experiments were performed in a dynamic gastrointestinal model (tiny-TIM) as described and validated for protein digestion previously (10,11,13,14). The model consists of a gastric compartment with a fundus and antrum function (12), a pyloric valve, and a small intestinal compartment. The food is mixed with added digestive enzymes, electrolytes, and bile by peristaltic movements of a flexible wall. The gastric content is gradually emptied into the small intestine via the pyloric valve. All settings are computer-controlled related to the age group to be simulated and the type of food. For dialysis of the digested products and for water absorption, a cartridge with hollow-fiber semi-permeable membranes (Sureflux-07L Hemodialyzer; Nipro, Belgium) was connected to the intestinal compartment. Dialysate was pumped through the hollow fibers at 10 mL/min. The dialyzed nutrients...
represent the compounds that are available for intestinal absorption (bioaccessible fraction) after digestion, such as small peptides and AA.

Simulated Infant Gastrointestinal Conditions

The test products were investigated in tiny-TIM simulating the conditions in the stomach and small intestine of healthy infants, 1 to 6 months of age, after the intake of IF (13). For this study, the following settings were used: gastric pH-curve from pH 6.7 to 4.8 from 0 to 30 minutes, to pH 3.8 from 30 to 60 minutes, and to pH 3.2 from 60 to 120 minutes, up to the end of the experiment by the addition of 1 mol/L HCl; gastric emptying curve with a halftime of 60 minutes and p-value of 1.5 (determines the s-shape of the emptying curve); intestinal pH at 6.5 ± 0.3 with 1 mol/L sodium bicarbonate.

Set-up of the In vitro Digestion Experiments

Each product and the blank were tested in duplicate for 4 hours (based on 3–4-hour feeding periods of infants). Blank experiments were performed to estimate the amount and digestibility of endogenous protein (enzyme and nonenzyme proteins from the pancreatin and bile). These blank experiments were performed with the intake of 118 g of water, 17 g of artificial saliva, and 12 g of citrate buffer.

Visual Observations

The gastric and intestinal walls of the compartments of the TIM systems are transparent, which enables visual observations during the digestion experiments. Every 10 minutes during the first hour of the experiments, the content in the gastric compartment was monitored and digital pictures were frequently made.

Sampling

During the experiments, dialysate samples were collected in 15- to 60-minute periods for 4 hours as follows: 0 to 15 minutes, 15 to 30 minutes, 30 to 45 minutes, 45 to 60 minutes, 60 to 90 minutes, 90 to 120 minutes, 120 to 180 minutes, and 180 to 240 minutes. The collected volume per period was measured and 3 subsamples of 30 mL were stored at −18°C for analysis.

Chemical Analysis

All collected dialysate samples from the duplicate tiny-TIM experiments were analyzed for total N (Kjeldahl) and α-amino nitrogen (AAN) (to calculate NH2-ending protein fragments). For AA analysis, the dialysate samples were pooled (0–4 hours). The Kjeldahl, AAN, and AA analysis were performed as described previously (11).

Data Analysis

The mean (±standard deviation; n = 2) bioaccessible amount of N and AA (dialyzed from the intestinal compartment) was calculated as absolute amount (volume × concentration). By subtracting the mean amount of N in the dialysate samples of the blank experiments (endogenous N) from that in the product samples (endogenous + exogenous N), the bioaccessible amount of exogenous N from the IF and HM was obtained for calculating the true ileal protein and AA digestibility as percentage of exogenous N or amino acid intake according to equation 1 (11): 

\[
\text{True ileal digestibility} [%] = \left( \frac{\text{Σ} \text{sample} [\text{mg}]}{\text{Σ} \text{blank} [\text{mg}]} \right) \times 100 \quad (1)
\]

The digestible IAA reference ratio of the IF and HM was calculated as described by the FAO (3) with equation 2:

\[
\text{DIAA reference ratio} = \frac{\text{digestible indispensable amino acid} [\text{mg}]}{\text{digestible indispensable amino acid} [\text{mg}]} \quad (2)
\]

The reference IAA scoring pattern for the DIAAS calculation for infants (0–6 months of age) is based on the IAA composition of HM. The DIAAS is the lowest calculated value of the DIAA reference ratio, expressed as percentage (3). Differences in true ileal protein and AA digestibility between GIF, CIF, and HM were statistically analyzed using analysis of variance followed by t tests using Tukey multiple comparisons test. Values were considered significantly different if \( P < 0.05 \).

RESULTS

Composition of Products

The analyzed N and AA composition of the reconstituted GIF and CIF were comparable. The measured N and calculated protein levels of HM were lower than those of the 2 reconstituted IFs (Table 1). The analyzed mean intake of N for the GIF, CIF, and HM was 340 ± 1 mg, 323 ± 1 mg, and 207 ± 1 mg, respectively. The total amount of N in the samples from the duplicate blank experiments was 280 ± 1 mg.

Visual Characteristics in the Gastric Compartment

Visual inspection of the milk products in the gastric compartment showed that for the CIF, the protein coagulation to particles was visible between 10 to 60 minutes after the start of the experiment. For the GIF, coagulation was visible around 10 to 30 minutes and for HM around 10 to 20 minutes.

True Ileal Digestibility and Digestion Kinetics of Proteins

The true ileal digestibility of the milk products, measured as amount of exogenous N in the dialysate collected per 15-, 30-, or 60-minute periods, is shown in Figure 1. The amount of bioaccessible exogenous N during the digestion of the GIF showed a similar kinetic profile as HM. In the first hour of digestion, the bioaccessible amount of N was 19.9% ± 3.5% and 23.3% ± 1.3% of N intake for the GIF and HM, respectively. The CIF showed a delay in the bioaccessible amount of N in the first hour of digestion: 11.2% ± 0.5% (\( P > 0.05 \) vs GIF; \( P < 0.05 \) vs HM) of intake. In the third hour of digestion, however, the bioaccessible amount of N was higher (\( P < 0.05 \) for the CIF (28.9% ± 1.2%) in comparison to the GIF (22.5% ± 1.6%) and HM (20.6% ± 1.0%).

The true ileal protein digestibility after 4 hours of digestion, expressed as percentage of N intake, showed no significant differences among the 3 test products: GIF 78.3% ± 3.7%, CIF 73.4% ± 2.7%, and HM 77.9% ± 4.1%.

The amounts of AAN expressed per gram N intake for each 1-h period is given in Figure 1. It shows the same pattern as the
bioaccessible N with lower amounts for CIF during the first 2 hours and higher amounts in the 3rd and 4th hour after intake in comparison to GIF and HM. The total values for AAN after 4 hours of digestion were 13.7, 13.1 and 14.0 mmol/g N for the GIF, CIF, and HM, respectively.

**True Ileal Digestibility of Indispensable Amino Acids and Digestible Indispensable Amino Acid Score**

The concentrations of all AAs (mg/100 mL product) in the reconstituted GIF and CIF and HM are given in Table 1. Based on the protein content in the milk products (corrected for non-protein N) and the data for the bioaccessible IAA in the 0- to 4-hour pooled dialysate samples, the true ileal digestibility of the IAA was calculated per gram of protein intake (Fig. 2). The DIAA reference ratio of each IAA for infants of 0 to 6 months of age is shown in Figure 3. The in vitro DIAAS for infants of the GIF, CIF, and HM, measured after 4 hours of digestion, was 83%, 75%, and 77%, respectively, with aromatic AA as first limiting AA for all 3 products. Protein quality with a DIAAS between 75% and 99% is regarded as ‘‘good’’ (3).

**DISCUSSION**

For determining the protein quality in human nutrition, the FAO recommends to determine the DIAAS, based on the true ileal protein and AA digestibility, preferably performed in humans (3). A suitable alternative is the use of in vitro dynamic, multicompart-mental gastrointestinal models simulating as optimal as possible the conditions in the upper gastrointestinal tract of infants. For simulation of infant conditions in tiny-TIM, 3 different settings were developed to simulate the most pronounced maturation stages of the gastrointestinal tract during the first period of life: neonates, infants, and toddlers, respectively. 0 to 1, 1 to 6, and 6 to 24 months of age (13). The FAO references have one age group for infants of 0 to 6 months of age. In the tiny-TIM experiments, the settings of 1- to 6-month-old infants were used, based on a range of physiological data (15,16) and corresponding with in vitro digestion of IF (17). In combination with previous validation experiments in tiny-TIM (9–11,14), we believe it gives reliable true ileal protein digestion data. To determine true ileal digestibility, the bioaccessibility of endogenous N was subtracted from that of the total N per time
In comparison to HM and the GIF, the protein digestion of the CIF was slower in the first hour, but faster in the third hour of digestion. Similar to HM, the bioaccessible N from the GIF could already be detected within 15 minutes after intake. It can be discussed whether this is related to differences in composition of the proteins with consequences for the physico-chemical reactions in the gastrointestinal tract. Differences in protein composition between goat and cow milk and HM are reported. For instance, goat milk contains more β-casein and less αs1-casein than cow milk, whereas β-casein is the major protein in HM in the absence of αs1-casein (6,25,26). A possible effect can be a difference in clotting behavior of casein in the stomach. The coagulates may consequently cause differences in gastric emptying of the protein because particles larger than a few millimeters will not easily pass the pyloric sphincter, in vivo as well as in tiny-TIM (27,28). Particles of clotted casein have to be broken down by pepsin before gastric emptying. Visual observation of the milk in the gastric compartment showed more persistent particles for the CIF compared to the GIF. For HM, less particles were visible for a shorter time period during gastric passage. Whether this shorter period of coagulation of the GIF versus CIF may influence gut physiology of infants needs further investigation. It has been suggested that in comparison to cow milk, goat milk forms softer and smaller curds in the stomach, which may facilitate digestive actions of the proteases (26). Another possible reason for early appearance of bioaccessible N in HM can be the differences in NPN content. As discussed above, HM contains a higher amount of NPN (20%–25%) than goat milk (15%) and cow milk (7%). This NPN fraction contains mainly free AA, urea, uric acid, and creatin (25). These compounds deliver relatively easily bioaccessible N. The appearance of bioaccessible AAN, however, shows a similar pattern as that of bioaccessible N, that is, higher amounts in the first 2 hours and lower amounts in the last 2 hours of digestion for HM and the GIF in comparison to the CIF. Therefore, the early appearance of bioaccessible N from HM and GIF might not come from the free AA, but from a faster digestion of the protein.

The bioaccessibility of the individual IAA per gram protein shows small differences between the CIF and HM, whereas the GIF showed somewhat higher levels for non-I AA.

The DIAAS for the GIF, CIF, and HM as determined for 0- to 6-month-old infants was 83%, 75%, and 77% for the aromatic AA when the protein intake was corrected for the nonamino acid nitrogen in the test products as discussed above. The FAO calculates with 25% NPN for HM (3), so, not corrected for free AA, resulting in a DIAAS for HM of 82%.

In the present study, the true ileal protein digestibility and DIAAS of the GIF and CIF are similar to those of HM. It can be concluded that the protein quality of the GIF and CIF, under simulated infant digestive conditions, is comparable to that of HM. The CIF shows a slower initial protein digestion in comparison to the GIF and HM. The underlying mechanisms can be investigated in future studies and whether 3 differences may have physiological consequences in infants after ingestion of IF.

Based on the accurate dynamic simulation of infant digestive conditions and the in vitro–in vivo correlation of protein digestion, we expect that the found DIAAS is reliable for infants. Owing to a lack of infant studies (because of ethical constraints), however, with intrinsically labeled proteins, a direct comparison of true AA digestibility in tiny-TIM (as well as in pigs) versus infants is not possible. Further discussions are necessary to evaluate whether dynamic gastrointestinal models are the best possible approach to determine protein quality for adults and infants given the restrictions of in vivo studies.
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